

A geometric method for model reduction of biochemical networks with polynomial rate functions

Satya Swarup Samal · Dima Grigoriev ·
Holger Fröhlich · Andreas Weber ·
Ovidiu Radulescu

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Abstract Model reduction of biochemical networks relies on the knowledge of slow and fast variables. We provide a geometric method, based on the Newton polytope, to identify slow variables of a biochemical network with polynomial rate functions. The gist of the method is the notion of tropical equilibration that provides approximate descriptions of slow invariant manifolds. Compared to extant numerical algorithms such as the intrinsic low dimensional manifold method, our approach is symbolic and utilizes orders of magnitude instead of precise values of the model parameters. Application of this method to a large collection of biochemical network models supports the idea that the number of dynamical variables in minimal models of cell physiology can be small, in spite of the large number of molecular regulatory actors.

Keywords Model Reduction · Algebraic Systems Biology · Complexity

Satya Swarup Samal

Algorithmic Bioinformatics, Bonn-Aachen International Center for IT, Dahlmannstraße 2, D-53113, Bonn, Germany E-mail: samal@cs.uni-bonn.de

Dima Grigoriev

CNRS, Mathématiques, Université de Lille, Villeneuve d'Ascq, 59655, France E-mail: dmitry.grigoryev@math.univ-lille1.fr

Holger Fröhlich

Algorithmic Bioinformatics, Bonn-Aachen International Center for IT, Dahlmannstraße 2, D-53113, Bonn, Germany E-mail: frohlich@bit.uni-bonn.de

Andreas Weber

Institut für Informatik II, University of Bonn, Friedrich-Ebert-Allee 144, 53113 Bonn, Germany E-mail: weber@cs.uni-bonn.de

Ovidiu Radulescu

DIMNP UMR CNRS 5235, University of Montpellier, Montpellier, France E-mail: ovidiu.radulescu@univ-montp2.fr

1 Introduction

Model reduction is an important problem in computational biology. There are several methods for reducing networks of biochemical reactions. Formal model reduction can be based on conservation laws, exact lumping [5], and more generally, symmetry [3,25]. Approximate numerical reduction methods, such as computational singular perturbation (CSP, [14]), intrinsic low dimensional manifold (ILDM, [17]) exploit the separation of timescales of various processes and variables. In dissipative systems, fast variables relax rapidly to some low dimensional attractive manifold called invariant manifold [7] that carries the slow mode dynamics. A projection of dynamical equations onto this manifold provides the reduced dynamics [17]. This simple picture can be complexified to cope with hierarchies of invariant manifolds and with phenomena such as transverse instability, excitability and itineracy. Firstly, the relaxation towards an attractor can have several stages, each with its own invariant manifold. During relaxation towards the attractor, invariant manifolds are usually embedded one into another (there is a decrease of dimensionality) [2]. Secondly, invariant manifolds can lose local stability, which allow the trajectories to perform large phase space excursions before returning in a different place on the same invariant manifold or on a different one [11]. The set of slow variables can change from one place to another. For all these reasons, even for fixed parameters, nonlinear models can have several reductions.

CSP and ILDM methods provide numerical approximations of the invariant manifold close to an attractor. These methods have been successfully applied to reduce networks of chemical reactions in chemical engineering. Other reduction methods utilize quasi-steady state or quasi-equilibrium approximations [9,22,23]. Numerical slow/fast decompositions result from the CSP or ILDM method, or from the calculation of a slowness index [22], in all cases relying on trajectory simulation. The application of these methods to computational biology is possible when model parameters are known. When parameters are unknown, or if they are known only by their orders of magnitudes, formal model reduction is needed. In addition, it is convenient to find reductions without having to simulate trajectories.

In this paper we propose a fully formal method to identify the slow and fast variables in a biochemical kinetic model with polynomial rate functions, without simulations of the trajectories. The method is based on computation of tropical equilibrations. Tropical methods, also known as max-plus algebras due their name to the fact that one of the pioneer of the field, Imre Simon, was Brazilian. These methods found numerous applications to computer science [29], physics [16], railway traffic [1], and statistics [21]. We have shown recently that they can be used to analyse systems of polynomial or rational differential equations with applications to cell cycle modelling [19]. The main idea of our approach is to identify situations when two or several monomials of different signs equilibrate each other and dominate all the remaining monomials in the right hand side of the differential equations defining the chemical kinetics. We call this situation tropical equilibration and show that it cor-

responds to partial equilibria of fast species [20]. Tropical equilibration was previously used in an interesting study by Savageau [28] as a design tool for network steady states. Our present focus is different because we are concerned with dynamics and model reduction. We also propose an algorithm using the Newton polytope to solve the tropical equilibration problem efficiently for large biochemical networks. An alternative algorithm for finding tropical equilibrations, based on constraint logic programming was proposed in [30]. However, when there are infinite branches of equilibrations, logic programming has no other alternative but the exhaustive enumeration of solutions between arbitrary bounds, whereas the Newton polytope method detects one solution per branch which is enough for identifying variable timescales and reduced models.

2 Approach

2.1 Dynamical equations and slow-fast decomposition

By slow-fast systems we understand dynamical systems for which some variables denoted by \mathbf{x} are fast and other variables denoted by \mathbf{y} are slow. Formally, the difference between slow and fast variables is indicated via a small positive parameter η such that:

$$\frac{d\mathbf{x}}{dt} = \frac{1}{\eta} \mathbf{f}(\mathbf{x}, \mathbf{y}) \quad (1)$$

$$\frac{d\mathbf{y}}{dt} = \mathbf{g}(\mathbf{x}, \mathbf{y}), \quad (2)$$

where f, g are functions not depending of η .

Let us suppose that the fast dynamics (1) has a stable state $\mathbf{x}^*(\mathbf{y})$ for all fixed \mathbf{y} values. Let $\mathbf{J}(\mathbf{y})$ be the linear operator (Jacobian) that gives the linearization of $\mathbf{f}(\mathbf{x}, \mathbf{y})$ at fixed \mathbf{y} , namely

$$\mathbf{f}(\mathbf{x}, \mathbf{y}) = \mathbf{J}(\mathbf{y})(\mathbf{x} - \mathbf{x}^*(\mathbf{y})) + O(|\mathbf{x} - \mathbf{x}^*(\mathbf{y})|^2).$$

We say that the stable state $\mathbf{x}^*(\mathbf{y})$ is uniformly hyperbolic if all the eigenvalues in the spectrum $Spec_{\mathbf{J}(\mathbf{y})}$ of $\mathbf{J}(\mathbf{y})$ have strictly negative real parts and are at a distance from the imaginary axis larger than $d > 0$, namely

$$Re(\lambda) < -d \text{ for all } \lambda \in Spec_{\mathbf{J}(\mathbf{y})} \text{ for all } \mathbf{y} \quad (3)$$

Tikhonov's theorem [33] says that if the above conditions are satisfied, then after a quick transition the system evolves approximately according to the following differential-algebraic equation:

$$\frac{d\mathbf{y}}{dt} = \mathbf{g}(\mathbf{x}, \mathbf{y}) \quad (4)$$

$$\mathbf{f}(\mathbf{x}, \mathbf{y}) = 0. \quad (5)$$

More precisely, the difference between solutions of (1),(2) and solutions of (4),(5) starting from the same initial data satisfying (5), vanishes asymptotically like a positive power of η when $\eta \rightarrow 0$.

Eq. (5) means that the fast variables are slaved by the slow ones. In this case, and given the condition (3) on the Jacobian of \mathbf{f} one can implicitly solve (5) and transform (4) into an autonomous reduced model for the slow variables. This approach is known as quasi-steady state approximation.

The first and most important step in the implementation of this reduction method is to find the slow-fast decomposition (1),(2), which means to identify \mathbf{x} , \mathbf{y} and η . For small models this can be done by rescaling variables and kinetic constants and by identifying the small parameter η as a ratio of kinetic constants or initial values of the variables. A well known example is the quasi-steady state approximation of the Michaelis-Menten enzymatic mechanism, when \mathbf{x} is the concentration of the enzyme-substrate complex, \mathbf{y} is the substrate concentration and η represents the ratio of the enzyme to the substrate plus product concentrations [20]. More generally, η can be interpreted as the ratio of fast to slow timescales. Numerical methods such as ILDM [17] use the Jacobian of the full system to obtain the slow-fast decomposition. In such methods η can be interpreted as the gap separating in logarithmic scale, the timescales of slow and fast variables obtained from the spectrum of the Jacobian. In this paper we present a symbolic method to perform the same decomposition. This method is based on tropical geometry.

2.2 Tropical equilibrations and timescales of the variables

We consider biochemical networks described by the following differential equations

$$\frac{dx_i}{dt} = \sum_{j \in [1, r]} k_j S_{ij} \mathbf{x}^{\alpha_j}, \quad i \in [1, n]. \quad (6)$$

where $k_j > 0, j \in [1, r]$ are kinetic constants, r is the number of reactions, S_{ij} are the elements of the so-called stoichiometric matrix, $\alpha_j = (\alpha_1^j, \dots, \alpha_n^j) \in \mathbb{Z}_+^n$ are multi-indices, $\mathbf{x}^{\alpha_j} = x_1^{\alpha_1^j} \dots x_n^{\alpha_n^j}$ and $x_i, i \in [1, n]$ are the species concentrations, n being the number of species.

The polynomial equations (6) can result from the mass action law. For instance, a reaction $A + B \rightarrow C$ of kinetic constant k and satisfying the mass action law, has $S_{11} = -1, S_{21} = -1, S_{31} = 1, \alpha_1 = (1, 1, 0)$, which correspond to the kinetic equations

$$\begin{aligned} \frac{dx_1}{dt} &= -kx_1x_2, \\ \frac{dx_2}{dt} &= -kx_1x_2, \\ \frac{dx_3}{dt} &= kx_1x_2, \end{aligned} \quad (7)$$

where x_1, x_2, x_3 are the concentrations of A, B, C , respectively.

However, the mass action law implies tight relations between α_j and S_{ij} , namely $\alpha_i^j = -S_{ij}$ if $S_{ij} < 0$, otherwise $\alpha_i^j = 0$. These relations are not used in our approach.

In what follows, the kinetic parameters do not have to be known precisely, they are given by their orders of magnitude. A convenient way to represent orders is by considering that

$$k_j = \bar{k}_j \varepsilon^{\gamma_j}, \quad \gamma_j = \text{round}(\log(k_j)/\log(\varepsilon)), \quad (8)$$

where ε is a positive parameter smaller than 1, \bar{k}_j has order unity. and round stands for the closest integer, with half-integers rounded to even numbers. This is the first step of the algorithm in Sect. 3.1.

We must emphasize that the parameter ε introduced in this section is not necessarily the fast/slow timescale ratio η occurring in Tikhonov's theorem. As a matter of fact, as will be shown later in this section, the parameter η can be expressed as a power of ε . In short, ε is used just for expressing everything as powers.

From (8) it follows that if $\gamma_j \neq \gamma_i$ and γ_i are integers, then $k_i/k_j > 1/\varepsilon$ or $k_j/k_i > 1/\varepsilon$, meaning that the parameters k_i, k_j are well separated. However, the condition $\gamma_j \neq \gamma_i$ is not always needed in this approach. All we need is the separation between the slow and fast timescales resulting from our calculations.

Timescales of nonlinear systems depend not only on parameters but also on species concentrations, which are a priori unknown. In order to compute them, we introduce a vector $\mathbf{a} = (a_1, \dots, a_n)$, such that $\mathbf{x} = \bar{\mathbf{x}}\varepsilon^{\mathbf{a}}$. Orders \mathbf{a} are unknown and have to be calculated. To this aim, the network dynamics can be described by a rescaled ODE system

$$\frac{d\bar{x}_i}{dt} = \left(\sum_j \varepsilon^{\mu_j} \bar{k}_j S_{ij} \bar{\mathbf{x}}^{\alpha_j} \right) \varepsilon^{-a_i}, \quad (9)$$

where $\mu_j = \gamma_j + \langle \mathbf{a}, \alpha_j \rangle$, and $\langle \cdot, \cdot \rangle$ stands for the vector dot product.

The r.h.s. of each equation in (9) is a sum of multivariate monomials in the concentrations. The exponents μ_j indicate how large are these monomials, in absolute value. Generically, one monomial of exponent μ_j dominates the others $\mu_j < \mu_{j'}, j' \neq j$. Accordingly, variables under the influence of a single dominant monomial, undergo large changes in a short time. The interesting case is when all variables are submitted to two dominant forces, one positive and one negative and these forces have the same order. We call this situation tropical equilibration ([20]) which corresponds to partial equilibration of fast variables on a slow invariant manifold. More precisely, we have the following

Definition 1 We call tropical equilibration solutions the vectors $\mathbf{a} \in \mathbb{R}^n$ where the minimum in the definition of the piecewise-affine function $f_i(\mathbf{a}) = \min_j (\gamma_j + \langle \mathbf{a}, \alpha_j \rangle)$ is attained at least twice for indices j', j'' corresponding to opposite signs monomials, i.e. $S_{ij'} S_{ij''} < 0$.

Intuitively, tropical equilibration means that dominant forces on variables compensate each other and that variables change slowly under the influence of the remaining weak forces. Compensation of dominant forces constrains the dynamics of the system to a low dimensional invariant manifold [19, 23, 20].

Tropical equilibrations are used to calculate the unknown orders \mathbf{a} . More precisely we want to solve the following system of min-plus equations:

$$\min_{j, S_{ij} < 0} (\gamma_j + \langle \mathbf{a}, \boldsymbol{\alpha}_j \rangle) = \min_{j', S_{ij'} > 0} (\gamma_{j'} + \langle \mathbf{a}, \boldsymbol{\alpha}_{j'} \rangle), \quad i = 1, \dots, n \quad (10)$$

The solutions of this system have a geometrical interpretation. Let us define the extended order vectors $\mathbf{a}^e = (1, \mathbf{a}) \in \mathbb{R}^{n+1}$ and extended exponent vectors $\boldsymbol{\alpha}_j^e = (\gamma_j, \boldsymbol{\alpha}_j) \in \mathbb{Z}^{n+1}$. Let us consider the equality $\mu_j = \mu_{j'}$. This represents the equation of a n dimensional hyperplane of \mathbb{R}^{n+1} , orthogonal to the vector $\boldsymbol{\alpha}_{j'}^e - \boldsymbol{\alpha}_j^e$:

$$\langle \mathbf{a}^e, \boldsymbol{\alpha}_{j'}^e \rangle = \langle \mathbf{a}^e, \boldsymbol{\alpha}_j^e \rangle, \quad (11)$$

where $\langle \cdot, \cdot \rangle$ is the dot product in \mathbb{R}^{n+1} . We will see in the next section that the minimality condition on the exponents μ_j implies that the normal vectors $\boldsymbol{\alpha}_{j'}^e - \boldsymbol{\alpha}_j^e$ are edges of the so-called Newton polytope [12, 31]. The algorithmic way to solve the set of inequalities in Eq. 10 along with the sign condition is described in Sects. 3.2, 3.3.

We call *tropically truncated system* the system obtained by pruning the system (9), i.e. by keeping only the dominating monomials.

$$\frac{d\bar{x}_i}{dt} = \varepsilon^{\nu_i} \left(\sum_{j \in J(i)} \bar{k}_j S_{ij} \bar{\mathbf{x}}^{\boldsymbol{\alpha}_j} \right), \quad (12)$$

where $J(i) = \underset{j}{\operatorname{argmin}} (\mu_j, S_{ij} \neq 0)$ selects the dominating rates of reactions acting on species i and

$$\nu_i = \min\{\mu_j | S_{ij} \neq 0\} - a_i \quad (13)$$

The tropically truncated equations contain generically two monomial terms of opposite signs (in special cases they can contain more than two terms). Polynomial systems with two monomial terms are called binomial or toric. In systems biology, toric systems are known as S-systems and were used by Savageau [27] for modeling metabolic networks.

The truncated system (12) indicates how fast is each variable, relatively to the others. The inverse timescale of a variable x_i is given by $\frac{1}{x_i} \frac{dx_i}{dt} = \frac{1}{\bar{x}_i} \frac{d\bar{x}_i}{dt}$ that scales like ε^{ν_i} . Thus, if $\nu_{i'} < \nu_i$ then $x_{i'}$ is faster than x_i .

Let us assume that the following gap condition is fulfilled:

$$\text{there is } f < n \text{ such that } \nu_{f+1} - \nu_f > 0, \quad (14)$$

meaning that two groups of variables have separated timescales. The variables $\mathbf{X}_f = (x_1, x_2, \dots, x_f)$ are fast (change significantly on timescales of order of magnitude $\varepsilon^{-\nu_f}$ or shorter). The remaining variables $\mathbf{X}_s = (x_{f+1}, x_{f+2}, \dots, x_n)$

are slow (have little variation on timescales of order of magnitude $\varepsilon^{-\nu_f}$). Then, the parameter $\eta = \varepsilon^{\nu_f+1-\nu_f}$ represents the fast/slow timescale ratio in the Tikhonov's theorem from the preceding section. Our gap condition means that η should be small. With these conditions, we have shown in [20,24] that quasi-steady state approximation can be applied. A further complication arises when the system has fast cycles and this will be described in the next section.

For systems with hierarchical relaxation, the separation between fast and slow variables is mobile within the cascade of relaxing modes. In the extreme case this means that all the variables timescales are separated by large enough gaps. Let us consider that we are interested in changes on timescales θ or slower. The timescale θ defines a threshold order value by the equation

$$\mu_{\text{threshold}} = -\log(\theta/\tau)/\log(\varepsilon), \quad (15)$$

where τ are the time units from the model. Then, from (12) it follows that all variables x_i with $\nu_i \geq \mu_{\text{threshold}}$ are slow. Perturbations in the concentrations of these species relax to an attractor slower or as slow as θ . The remaining species are fast and the perturbations in their concentrations relax to equilibrated values much faster than θ .

2.3 Model reduction of fast cycles

Tropical truncation is useful for identifying the slow and fast variables of a system of polynomial differential equations. However, the truncation alone is not always enough for accurate reduction. As discussed in [20,24], there are situations when the truncated system is not a good approximation. Typically, truncation could eliminate all the reactions exiting a fast cyclic subnetwork. Thus we get new conserved quantities, that were not conserved by the full model. Truncation is in this case accurate at short times, but introduce errors at large times. In order to cope with fast cycles pruning, we adopt the recipe discussed in [9] for the quasi-equilibrium approximation. This recipe allows to recover the terms that were neglected by truncation, but which are important for large time dynamics.

First, let us remind some definitions. We call linear conservation law of a system of differential equations, a linear form $C(\mathbf{x}) = \langle \mathbf{c}, \mathbf{x} \rangle = c_1x_1 + c_2x_2 + \dots + c_nx_n$ that is identically constant on trajectories of the system. It can be easily checked that vectors in the left kernel $\text{Ker}^l(S)$ of the stoichiometric matrix \mathbf{S} provide linear conservation laws of the system (6). Indeed, system (6) reads $\frac{d\mathbf{x}}{dt} = \mathbf{S}\mathbf{R}(\mathbf{x})$, where the components of the vector \mathbf{R} are $R_j(\mathbf{x}) = k_jx^{\alpha_j}$. If $\mathbf{c}\mathbf{S} = 0$, then $\frac{d\langle \mathbf{c}, \mathbf{x} \rangle}{dt} = \mathbf{c}\mathbf{S}\mathbf{R}(\mathbf{x}) = 0$, where $\mathbf{c} = (c_1, c_2, \dots, c_n)$.

Let us assume that the truncated system (12), restricted to the fast variables has a number of independent, linear conservation laws, defined by the left kernel vectors $\mathbf{c}_1, \mathbf{c}_2, \dots, \mathbf{c}_d$, where $\mathbf{c}_k = (c_{k1}, c_{k2}, \dots, c_{kf})$. These conservation laws can be calculated by recasting the truncated system as the product of a new stoichiometric matrix and a vector of monomial rate functions and further computing left kernel vectors of the new stoichiometric matrix. We

further assume that the fast conservation laws are not conserved by the full system (6).

We define some new slow variables $\mathbf{Y} = (y_1, \dots, y_d)$, where $y_k = \sum_{i=1}^f c_{ki}x_i$ and eliminate the fast variables x_1, x_2, \dots, x_f by using the system :

$$\sum_{j \in J(i)} k_j S_{ij} \mathbf{x}^{\alpha_j} = 0, \quad i \in [1, f] \quad (16)$$

$$\sum_{i=1}^f c_{ki} x_i = y_k, \quad k \in [1, d] \quad (17)$$

Reactions of the initial model that were pruned by truncation have to be restored if they act on the new slow variables \mathbf{Y} , i.e. if $\sum_{i=1}^f c_{li} S_{ik} \neq 0$, for some $l \in [1, d]$, where k is the index of the reaction to be tested. Finally, the kinetic laws of these reactions have to be redefined in terms of the slow variables \mathbf{X}^s, \mathbf{Y} .

The rigorous justification of the reduction procedure for models with fast cycles can be found in [24].

3 Algorithm to compute tropical equilibrations.

In this section we introduce an algorithm allowing the automatic computation of tropical equilibrations.

3.1 Pre-processing

We consider examples with polynomial vector field. The kinetic parameters of the equation system are scaled based on Eq. (8).

3.2 Newton polytope and edge filtering

For each equation and species i , we define a Newton polytope \mathcal{N}_i , that is the convex hull of the set of points α_j^e such that $S_{ij} \neq 0$ and also including together with all the points the half-line emanating from these points in the positive ε direction. This is the Newton polytope of the polynomial in right hand side of Eq. (9), with the scaling parameter ε considered as a new variable.

As explained in Sect. 2.2 the tropical equilibrations correspond to vectors $\mathbf{a}^e = (1, \mathbf{a}) \in \mathbb{R}^{n+1}$ satisfying the optimality condition of Definition 1. This condition is satisfied automatically on hyperplanes orthogonal to edges of Newton polytope connecting vertices $\alpha_{j'}^e, \alpha_{j''}^e$ satisfying the opposite sign condition. Therefore, a subset of edges from Newton polytope is selected based

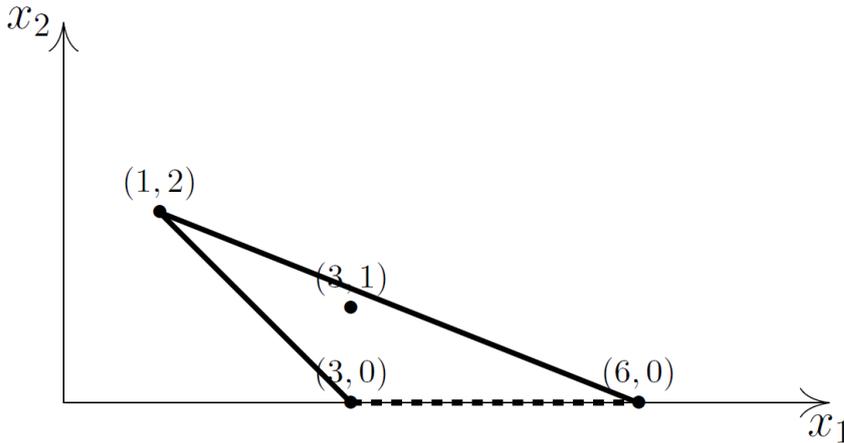


Fig. 1 An example of a Newton polytope for the polynomial $-x_1^6 + x_1^3x_2 - x_1^3 + x_1x_2^2$. In this example, all the monomial coefficients have order zero in ε and we want to solve the tropical problem $\min(3a_1 + a_2, a_1 + 2a_2) = \min(6a_1, 3a_1)$. The Newton polytope vertices $(6, 0), (3, 0), (1, 2)$ are connected by lines. The point $(3, 1)$ is not a vertex as it lies in the interior of the polytope. This stems to having $\min(3a_1 + a_2, a_1 + 2a_2) = a_1 + 2a_2$ for all tropical solutions, which reduces the number of cases to be tested. The thick edges satisfy the sign condition, whereas the dashed edge does not satisfy this condition. For this example, the solutions of the tropical problem are in infinite number and are carried by the two half-lines $a_1 = a_2 \geq 0$ and $5a_1 = 2a_2 \leq 0$, orthogonal to the thick edges of the Newton polygon.

on the filtering criteria which tells that the vertices belonging to an edge should be from opposite sign monomials as explained in Eq. (18).

$$E(P) = \{\{v_1, v_2\} \subseteq \binom{V}{2} \mid \text{conv}(v_1, v_2) \in F_1(P) \wedge \text{sign}(v_1) \times \text{sign}(v_2) = -1\}, \quad (18)$$

where v_i is the vertex of the polytope and V is the vertex set of the polytope, $\text{conv}(v_1, v_2)$ is the convex hull of vertices v_1, v_2 and $F_1(P)$ is the set of 1-dimensional face (edges) of the polytope. $\text{sign}(v_i)$ represents the sign of the monomial which corresponds to vertex v_i . Fig. 1 shows an example of Newton polytope construction for a single equation. Further definitions about properties of a polytope and Newton polytope can be found in [12, 31].

3.3 Pruning and feasible solutions

By feasible solution we understand a vector (a_1, \dots, a_n) satisfying all the equations of the system (10). A feasible solution lies in the intersection of hyperplanes (or convex subsets of these hyperplanes) orthogonal to edges of Newton polytopes obeying the sign conditions. Of course, not all sequences of edges lead to nonempty intersections and thus feasible solutions. This can be tested

by the following linear programming problem, resulting from (10):

$$\begin{aligned} \gamma_j(i) + \langle \mathbf{a}, \boldsymbol{\alpha}_j(i) \rangle &= \gamma'_j(i) + \langle \mathbf{a}, \boldsymbol{\alpha}'_j(i) \rangle \leq \gamma''_j + \langle \mathbf{a}, \boldsymbol{\alpha}''_j \rangle, \\ &\text{for all } j'' \neq j, j', S_{ij''} \neq 0, \quad i = 1, \dots, n \end{aligned} \quad (19)$$

where $j(i), j'(i)$ define the chosen edge of the i -th Newton polytope. The set of indices j'' can be restricted to vertices of the Newton polytope, because the inequalities are automatically fulfilled for monomials that are internal to the Newton polytope. For instance, in the example of the preceding section, the choice of the edge connecting vertices (1, 2) and (6, 0) leads to the following linear programming problem:

$$a_1 + 2a_2 = 3a_1 \leq 6a_1,$$

whose solution is a half-line orthogonal to the edge of the Newton polygon.

We introduce a pruning methodology (similar to a branch and bound algorithm technique) which helps to reduce the number of possible choices of Newton polytope edges leading to feasible solutions. Let us consider a system of polynomial equations and order the equations as eq_1, eq_2, \dots, eq_n . Let the vertices of Newton polytope \mathcal{N}_n be $v_{n1}, v_{n2}, \dots, v_{nl}$ where l is the total number of vertices. The polytope edges are described by ne_1, ne_2, \dots, ne_n where ne_i denotes the set of edges from Newton polytope \mathcal{N}_i . In order to search for feasible solutions an edge from each polytope needs to be selected. This translates to evaluating the cartesian product of ne_1, ne_2, \dots, ne_n which can be described by the following equation

$$\begin{aligned} ne_1 \times ne_2 \times \dots \times ne_n &= \{(e_{1j}, e_{2j}, \dots, e_{nj}) \mid (e_{1j} \\ &\in ne_1) \wedge (e_{2j} \in ne_2) \wedge \dots \wedge (e_{nj} \in ne_n)\}. \end{aligned} \quad (20)$$

where e_{nj} is the j^{th} edge in Newton polytope \mathcal{N}_n . It is clear from the above that the possible choices are exponential. In order to improve the running time of the algorithm, the pruning strategy evaluates Eq. (20) in several steps (cf. Algorithm 1 and Fig. 2). It starts with an arbitrary pair of edges and proceeds to add the next edge only when the inequalities (19) restricted to these two pair of edges are satisfied. The corresponding set of inequalities can be solved using any standard linear programming package.

3.4 Example

As an illustration of our method we wanted a simple model that (i) has polynomial dynamics and (ii) contains fast cycles that ask for the reduction steps described in Sect.2.3. A cell cycle model proposed by Tyson [34] satisfies both these conditions. This model describes the interaction between cyclin and cyclin-dependent kinase *cdc2* during the progression of the eukaryotic cell cycle (see Fig. 3). Cyclin (variable x_5) is synthesized during interphase stage of the cycle (reaction of constant k_6). Newly synthesized cyclin forms a complex with

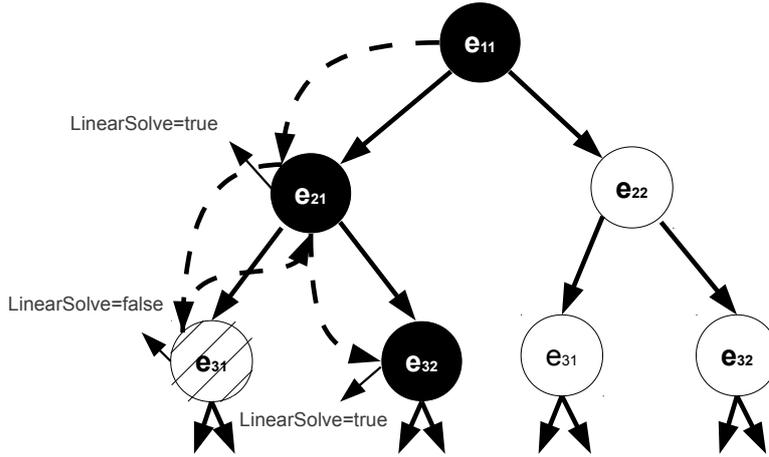


Fig. 2 Pruning strategy. This figure explains the pruning technique described in Sect. 3.3 and the evaluation of Eq. (20). The possible combinations of edges are represented in a tree representation. The algorithm starts by testing for feasible solution for first pair of edge sets. If a feasible solution is found, the algorithm proceeds further to other edge sets or it backtracks. In the figure, e_{11} and e_{21} are selected from edge sets ne_1, ne_2 and are checked for a feasible solution satisfying (19). If such a solution exists, it moves to e_{31} from the next edge set and again checks for feasible solution, if not then it backtracks to e_{21} and then to e_{32} which results in a feasible solution. Therefore, the sub-tree with root node e_{31} is discarded from future searches and this improves running time. Likewise the branch e_{11} and e_{22} is explored. This approach is similar to branch and bound algorithm technique. The dashed arrows show the flow of the program.

Algorithm 1: SolveOrders: Steps of tropical equilibration algorithm

Input: List of edge sets ne_1, ne_2, \dots, ne_n , and the corresponding vertices
Output: Orders of the variables a_1, a_2, \dots, a_n (tropical equilibration solution set)

```

1 begin
2   solutionset = {}; integer k=1; equation = {}
3   SolveOrders(equation, k, edge-sets, vertices)
4   if k > n then
5     return
6   for l = 1 to number of entries in ne_k edge-set do
7     equation(k)* = vertices in lth row
8     inequalities* = all other vertices in ne_1 to ne_k edge-sets
9     if LinearSolve(equation, inequalities) is feasible then
10      if k = n then
11        add the solution of LinearSolve to solutionset
12      SolveOrders(equation, k + 1, ne_1, ..., ne_k, vertices)

```

13 *The equations and inequalities are initialised as per Eq. (19)

the phosphorylated kinase *cdc2* (*cdc2* is the variable x_2 and the complex formation reaction has constant k_4). The resulting complex (variable x_4) is called inactive or pre-maturation promoter factor (pre-MPF). pre-MPF needs to be activated for enter into mitosis in order to phosphorylate many substrates controlling processes essential for nuclear and cellular division. The active form of MPF (variable x_3) is produced from pre-MPF either by a non-regulated transformation (reaction of constant k_{10}) or by an autocatalytic process (reaction of constant k_9). At the end of mitosis the active complex dissociates (reaction of constant k_1), resulting in the phosphorylated cyclin (variable x_6) that is degraded (reaction of constant k_8) and the de-phosphorylated kinase *cdc2* (variable x_1). The kinase is equilibrated with its phosphorylated form (variable x_2) by phosphorylation and dephosphorylation reactions (of constants k_2 and k_3 respectively).

The full model has a stable periodic attractor, a limit cycle. The stable limit cycle oscillations correspond to the periodic succession of interphase and mitosis phases of the cell cycle.

The corresponding system of differential equations along with conservation laws for the above model can be described as

$$\begin{aligned}
\dot{x}_1 &= k_1x_3 - k_2x_1 + k_3x_2, & \dot{x}_2 &= k_2x_1 - k_3x_2 - k_4x_2x_5, \\
\dot{x}_3 &= k_{10}x_4 - k_1x_3 + k_9x_3^2x_4, \\
\dot{x}_4 &= k_4x_2x_5 - k_{10}x_4 - k_9x_3^2x_4, & \dot{x}_5 &= k_6 - k_4x_2x_5, \\
\dot{x}_6 &= k_1x_3 - k_8x_6, & x_1 + x_2 + x_3 + x_4 &= 1
\end{aligned} \tag{21}$$

For this model, the tropical equilibration problem reads :

$$\begin{aligned}
\min(a_3 + \gamma_1, a_2 + \gamma_3) &= a_1 + \gamma_2, \\
a_1 + \gamma_2 &= \min(a_2 + \gamma_3, a_2 + a_5 + \gamma_4), \\
\min(a_4 + \gamma_{10}, 2a_3 + a_4 + \gamma_9) &= a_3 + \gamma_1, \\
a_2 + a_5 + \gamma_4 &= \min(a_4 + \gamma_{10}, 2a_3 + a_4 + \gamma_9), \\
\gamma_6 &= a_2 + a_5 + \gamma_4, \\
a_3 + \gamma_1 &= a_6 + \gamma_8, \min(a_1, a_2, a_3, a_4) = 0
\end{aligned} \tag{22}$$

Using the numerical values of the parameters from the original paper we find, for $\varepsilon = 1/9$, $\gamma_1 = 0$, $\gamma_2 = -6$, $\gamma_3 = -3$, $\gamma_4 = -2$, $\gamma_6 = 2$, $\gamma_8 = 0$, $\gamma_9 = -2$, $\gamma_{10} = 2$ (cf. Eq. 8 and Sect. 3.1).

Remark: One may notice that the orders γ depend on which units were used for the parameters. However, if the parameter units are changed, the set of tropical equilibrations is transformed into an equivalent one. Indeed, the model equations should be invariant with respect to units conversion. In particular, if units of second order reaction constants (i.e. coefficients of second order monomial rates) are multiplied by k , one should subtract $\log(k)/\log(\varepsilon)$ from the parameter orders and add the same quantity to the concentration orders. This will generate an equivalent set of solutions, up to rounding errors.

Using our algorithm (cf. Sects.3.2, 3.3) we got three tropical equilibrations for this system, namely $\mathbf{a}_1 = (8, 5, 2, 0, -1, 2)$, $\mathbf{a}_2 = (5, 2, 2, 0, 2, 2)$, $\mathbf{a}_3 = (3, 0, 2, 0, 4, 2)$.

The rescaled truncated system for the solution \mathbf{a}_3 reads

$$\begin{aligned}\dot{\bar{x}}_1 &= \varepsilon^{-6}(\bar{k}_3\bar{x}_2 - \bar{k}_2\bar{x}_1), \dot{\bar{x}}_2 = \varepsilon^{-3}(\bar{k}_2\bar{x}_1 - \bar{k}_3\bar{x}_2), \\ \dot{\bar{x}}_3 &= \bar{k}_{10}\bar{x}_4 - \bar{k}_1\bar{x}_3 + \bar{k}_9\bar{x}_3^2\bar{x}_4, \\ \dot{\bar{x}}_4 &= \varepsilon^2(-\bar{k}_{10}\bar{x}_4 + \bar{k}_4\bar{x}_2\bar{x}_5 - \bar{k}_9\bar{x}_3^2\bar{x}_4), \\ \dot{\bar{x}}_5 &= \varepsilon^{-2}(\bar{k}_6 - \bar{k}_4\bar{x}_2\bar{x}_5), \dot{\bar{x}}_6 = \bar{k}_1\bar{x}_3 - \bar{k}_8\bar{x}_6.\end{aligned}\quad (23)$$

It appears clearly that the variables x_1, x_2, x_5 are fast. More precisely, their characteristic times are $\nu_1^{-1} = \varepsilon^6$, $\nu_2^{-1} = \varepsilon^3$, $\nu_5^{-1} = \varepsilon^2$, respectively. The largest of these timescales is here approximately 0.01 (in minutes which are the time units of the model). The remaining slow variables have characteristic times from ε^0 to ε^{-2} , i.e. approximately from 1 to 100 min. Therefore, the timescales of slow and fast species are separated by a gap, and the singular perturbation small parameter (cf. Sect.2.1) is $\eta = t_{fast}/t_{slow} \sim \varepsilon^2$ (the power 2 arises as the difference between $\nu_6 = \nu_3 = 0$, coming from the fastest slow species and $\nu_5 = -2$, coming from the slowest fast species).

The fast truncated system reads

$$\begin{aligned}\dot{x}_1 &= k_3x_2 - k_2x_1, \dot{x}_2 = k_2x_1 - k_3x_2, \\ \dot{x}_5 &= k_6 - k_4x_2x_5\end{aligned}\quad (24)$$

and has a single conservation law $C_1 = x_1 + x_2$ that provides a new slow variable. This conservation law, not conserved by the full system (21), indicates the presence of a fast cycle in the model. It is the rapid phosphorylation/dephosphorylation cycle transforming the cyclin x_1 into its phosphorylated form x_2 and back. The fast variables are eliminated from the system obtained by adding to (24) the definition of the fast conservation law cf. Sect.2.3:

$$k_3x_2 - k_2x_1 = 0, k_6 - k_4x_2x_5 = 0, y = x_1 + x_2. \quad (25)$$

The differential equation for y is obtained by adding the first two equations of the full system (21), and thus restoring the terms k_1x_3 and $k_4x_2x_5$, that have order ε^2 and were pruned in the first step.

Finally, we obtain the following reduced model

$$\dot{x}_3 = k_{10}x_4 - k_1x_3 + k_9x_3^2x_4, \quad (26)$$

$$\dot{x}_4 = -k_{10}x_4 + k_6 - k_9x_3^2x_4, \quad (27)$$

$$\dot{x}_6 = k_1x_3 - k_8x_6, \dot{y} = k_1x_3 - k_6. \quad (28)$$

and the slaved fast variables are given by $x_1 = yk_2/(k_2 + k_3) \approx yk_2/k_3$, $x_2 = yk_3/(k_2 + k_3) \approx y$, $x_5 = k_6(k_2 + k_3)/(k_4k_3y)$, where we have used the fact that $k_2 \ll k_3$.

Let us note that the variable y has the same order as x_2 ($a_y = a_3 = 2$), it is tropically equilibrated ($\gamma_1 + a_3 = \gamma_6 = 2$ in Eq.(28)), and has $\nu_y = \gamma_6 - a_y = 0$ meaning that it is slow.

Let us call this four variables model Reduced Model 1. Note that in this model the dynamics of the variables x_3, x_4 is decoupled from the two others. We can therefore conclude that by our approach we obtain a two dimensional minimal cell cycle model.

Repeating the procedure for the equilibrations $\mathbf{a}_1, \mathbf{a}_2$ we find two other rescaled truncated systems.

The rescaled truncated system for the solution \mathbf{a}_1 reads

$$\begin{aligned}\dot{\bar{x}}_1 &= \varepsilon^{-6}(\bar{k}_3\bar{x}_2 - \bar{k}_2\bar{x}_1 + k_1x_3), \dot{\bar{x}}_2 = \varepsilon^{-3}(\bar{k}_2\bar{x}_1 - \bar{k}_3\bar{x}_2), \\ \dot{\bar{x}}_3 &= \bar{k}_{10}\bar{x}_4 - \bar{k}_1\bar{x}_3 + \bar{k}_9\bar{x}_3^2\bar{x}_4, \\ \dot{\bar{x}}_4 &= \varepsilon^2(-\bar{k}_{10}\bar{x}_4 + \bar{k}_4\bar{x}_2\bar{x}_5 - \bar{k}_9\bar{x}_3^2\bar{x}_4), \\ \dot{\bar{x}}_5 &= \varepsilon^3(\bar{k}_6 - \bar{k}_4\bar{x}_2\bar{x}_5), \dot{\bar{x}}_6 = \bar{k}_1\bar{x}_3 - \bar{k}_8\bar{x}_6,\end{aligned}\quad (29)$$

and for the solution \mathbf{a}_2 we got

$$\begin{aligned}\dot{\bar{x}}_1 &= \varepsilon^{-6}(\bar{k}_3\bar{x}_2 - \bar{k}_2\bar{x}_1), \dot{\bar{x}}_2 = \varepsilon^{-3}(\bar{k}_2\bar{x}_1 - \bar{k}_3\bar{x}_2), \\ \dot{\bar{x}}_3 &= \bar{k}_{10}\bar{x}_4 - \bar{k}_1\bar{x}_3 + \bar{k}_9\bar{x}_3^2\bar{x}_4, \\ \dot{\bar{x}}_4 &= \varepsilon^2(-\bar{k}_{10}\bar{x}_4 + \bar{k}_4\bar{x}_2\bar{x}_5 - \bar{k}_9\bar{x}_3^2\bar{x}_4), \\ \dot{\bar{x}}_5 &= (\bar{k}_6 - \bar{k}_4\bar{x}_2\bar{x}_5), \dot{\bar{x}}_6 = \bar{k}_1\bar{x}_3 - \bar{k}_8\bar{x}_6.\end{aligned}\quad (30)$$

In both cases, the variable x_5 is slow, which was not the case for the equilibration \mathbf{a}_1 . This is possible, because for a nonlinear model, the timescale of a variable depends on the concentration range in which the model functions. The equilibrations \mathbf{a}_1 and \mathbf{a}_2 correspond to low and very low concentrations of phosphorylated kinase x_2 (proportional to ε^5 and ε^2 , respectively), meaning slow consumption of the cyclin x_5 . The concentration of x_2 is large for the equilibration \mathbf{a}_3 (proportional to ε^0) leading to rapid consumption of x_5 (see Eq.(21)).

The two tropical equilibrations \mathbf{a}_1 and \mathbf{a}_2 lead to the same reduced model, which we call Reduced model 2:

$$\begin{aligned}\dot{x}_3 &= k_{10}x_4 - k_1x_3 + k_9x_3^2x_4, \\ \dot{x}_4 &= -k_{10}x_4 + k_6 - k_9x_3^2x_4, \\ \dot{x}_5 &= k_6 - k_4yx_5, \\ \dot{x}_6 &= k_1x_3 - k_8x_6, \dot{y} = k_1x_3 - k_6,\end{aligned}\quad (31)$$

to be considered together with $x_1 = yk_2/k_3, x_2 = y$.

The tropical setting confirms ideas from the theory of nonlinear dynamical systems. The two reduced models are nested. Reduced model 2 has a larger number of slow (relaxing) variables than Reduced model 1. This means that the corresponding invariant manifolds are embedded one into another with the lowest dimensional one defined by the Reduced model 1 carrying the dynamics on the limit cycle attractor. Starting with initial low concentrations of the phosphorylated kinase corresponding to the equilibration \mathbf{a}_1 or \mathbf{a}_2 , the system will increase these concentrations to levels corresponding to the equilibration \mathbf{a}_3 that allow the stable limit cycle oscillations.

One can notice that our Reduced model 1 does not contain the parameters k_2, k_3, k_4 of the full model. This means that as long as the phosphorylation and the dephosphorylation of the free kinase, as well as the formation of cyclin kinase complex are fast enough, the actual values of the kinetic constants of these processes are not important. This proves the robustness of the full model with respect to changes of these parameters: these parameters are insensible. More generally, as discussed elsewhere [23], model reduction can be used to classify the parameters of a model as sensible and insensible.

In his paper, Tyson [34] also proposes a two variables reduced model:

$$\begin{aligned}\dot{u} &= k_9(v - u)(\alpha + u^2) - k_1u, \\ \dot{v} &= k_6 - k_1u,\end{aligned}\tag{32}$$

where $u = x_3$, $v = x_3 + x_4 + x_5$, $\alpha = k_{10}/k_9$.

It can be easily checked that Eqs.(32) are equivalent with our Eqs.(26),(27), provided that $x_5 \ll x_4$ and $x_5 \ll x_3$. These last conditions, justified by intuitive arguments, were used in the derivation of the reduced model in [34]. In our approach, the same conditions follow immediately from the orders of the species concentrations. Indeed, for the equilibration \mathbf{a}_3 we have $x_5 \sim \varepsilon^4$, $x_3 \sim \varepsilon^2$, $x_4 \sim \varepsilon^0$, therefore $x_5 \ll x_4$ and $x_5 \ll x_3$. To summarize, the advantage of our approach is that it is automatic and can be applied to larger models that are more difficult or impossible to grasp by simple intuition.

4 Results

4.1 Details on the implementation of the algorithm

The differential equation system along with the conservation laws for a given biochemical model (in SBML format) are generated using the pocab software[26]. Polymake [6] is used to compute the Newton polytope from the set of exponent vectors. For solving the linear programming we used Gurobi[10] in Java programming environment. The expressions in the equation system are processed using computer algebra system Maple.¹

4.2 Results on Biomodels database

Biological models were selected from r25 version of Biomodels [15]. For our analysis, we selected models with polynomial kinetics. We performed two analysis. (i) First, we benchmarked our implementation against models derived from Biomodels. (ii) Second, we computed the average number of slow variables across Biomodels with different time thresholds to get an estimate of the dimension of the invariant manifold.

¹ The code can be downloaded together with supporting information from <http://www.abi.bit.uni-bonn.de/index.php?id=17>.

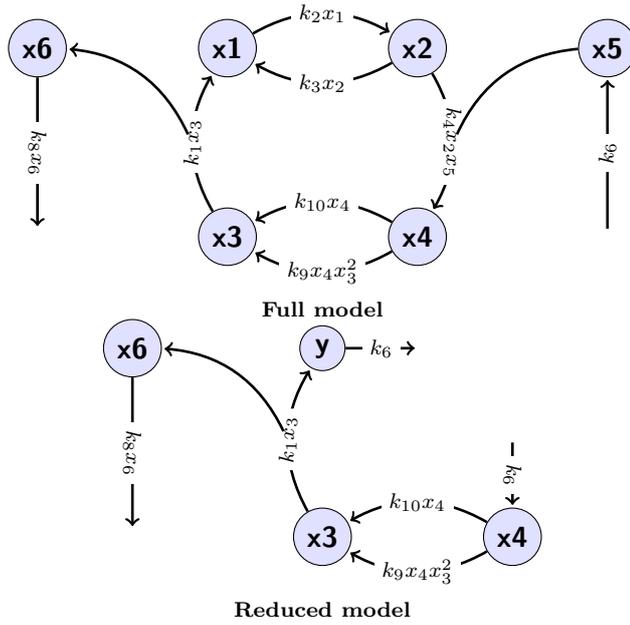


Fig. 3 Graphic representation of the full cell cycle model [34] (referenced as BIOMD00000005 by Biomodels) and of the reduced model 1. The full model describes the cyclin production and complex formation between the cyclin and the kinase cdc2, the auto-catalytic activation (by dephosphorylation), and the dissociation of this complex followed by the destruction of the cyclin. The reduced model represents accurately the same processes, on the invariant manifold containing the periodic attractor. The different variables mean: x_1 : cdc2, x_2 : cdc2-P, x_3 : cdc2 : cyclin-P i.e. active MPF complex, x_4 : P-cdc2 : cyclin-P i.e. pre-MPF complex, x_5 : cyclin, x_6 : cyclin-P, $y = x_1 + x_2$: total free cdc2. Both full and reduced model are biochemical networks with polynomial rate functions.

In the first analysis, the parameters were replaced by the orders of magnitude according to Eq. (8). A summary of the analysis is presented in Table 1. The analysis is performed to compute all possible combinations of vertices leading to tropical solutions within a maximal running time of 10000 seconds of CPU time. The CPU time threshold for models with ε value = $1/5$ was further increased to 100,000 to further ascertain the exponential behaviour. In practice, we restrict this search space using the tree pruning strategy as explained in Sect. 3.3. While solving the linear inequalities it may happen that there exists infinite feasible solutions for a given combination of vertices. In such a scenario we report only one solution. The analysis was repeated with four different values for ε . A semilog time-plot is presented in Fig. 4 which plots the log of running time in milliseconds versus the number of equations in the model. It should be pointed out that the number of variables may not be equal to the number of equations because of conservation laws which were treated as extra linear equations in our framework.

In the second analysis, the number of slow variables were computed based on the rescaled orders i.e. $\mu_i - a_i > \mu_{\text{threshold}}$ and a certain time threshold as explained in Eq. (15). In addition to slow species, we computed the quasi buffered species which are slow variables with very high time threshold. To compute them we fixed the timescale threshold to 100,000 seconds and the slow species at this threshold are labeled as quasi buffered species. In the model, such species are practically constant and in our setting these are subtracted from slow variables. A boxplot is shown in Fig. 5 where a point represents the compression ratio (i.e. ratio of average number of slow variables / total number of variables) over all the tropical equilibrations for each model with respect to different time thresholds. This was performed for all the models under consideration. We also computed the slowest timescale for each model which is defined as the smallest time threshold at which all species become fast (the quasi-buffered species were removed from the model before performing this step). A histogram showing the distribution slowest timescale is presented in Fig. 5. To estimate the slowest timescale, the time threshold is varied and the number of slow species are counted, the threshold at which all species become fast is considered to be the slowest timescale of that model. This histogram indicates that the benchmarked models are representative of a wide variety of cellular processes whose timescales range from fractions of seconds to one day.

The calculations in this section were performed for different values of the parameter ε . According to the geometric interpretation of tropical equilibrations in Sect.2.2 the tropical solutions are either isolated points or bounded or unbounded polyhedra. Changing the parameter ε is just a way to approximate the position of these points and polyhedra by lattices or in other words by integer coefficients vectors. The approximation results from the rounding in Eq.(8) and better approximations would be to consider rational (with a largest common denominator), instead of integer orders γ_i and a_i . Finding the value of ε that provides the best approximation is a complicated problem in Diophantine approximation. For that reason, we preferred an experimental approach consisting in choosing several values of ε and checking the robustness of the results.

4.3 Tree Pruning

In order to evaluate the efficacy of tree pruning, we computed the ratio between number of times the linear programming is invoked with every tree pruning step (cf. Fig. 2) and the possible number of combinations of Newton polytope edges without tree pruning (cf. Eq. 20). This ratio is a measure of efficiency achieved due to pruning. From the computations, it can be seen that for smaller dimensional models the log of this ratio is greater than 0 meaning tree pruning works worse by invoking linear programming more than what is required without tree pruning. However, for the majority of large dimensional models the log of this ratio is less than 0 suggesting significant reduction in

Table 1 Summary of analysis on Biomodels database. Tropical solutions here mean existence of at least one feasible solution from all possible combination of vertices of the Newton polytope (in case of infinite solutions, one is picked (cf. Sect. 4.2)). Timed-out means all solutions could not be computed within 10,000 seconds of computation time (except for models with ε value = 1/5). No tropical solution implies no possible combination of vertices could be found resulting in a feasible solution. Unit-definition refers to the presence of SBML tag <unitDefinition> which defines the time units of the model to be used in Eq. (15). For the models where it is absent seconds is taken to be the default time unit of the model, with the exception of two models whose units have been curated manually by comparison with original papers.

ε value	Total models considered	Timed-out models	Models without tropical solutions	Models with tropical solutions	Average running time (in secs)**	Models with Unit-definition
1/5*	53	14	0	39	1354.91	12
1/7	53	17	0	36	512.07	12
1/9	53	17	0	36	432.54	12
1/11	53	16	0	37	756.66	12
1/19	53	18	2	33	1063.30	12
1/23	53	18	1	34	783.98	12
1/47	53	18	0	35	719.79	12
1/53	53	19	0	34	387.16	12
1/59	53	19	1	33	482.30	12
1/71	53	19	0	34	640.32	12

*For this ε the running time threshold was 100,000 secs. ** For average time computation, the running times of those models which did not timed-out (i.e. 4th and 5th column) were considered.

the search space due to pruning. The results for ε value of 1/5 are presented in Fig. 4.

4.4 Testing the method

In order to test the method we consider the cell cycle model [34] (referenced as BIOMD00000005 by Biomodels) in more detail. This is the cell cycle model example analysed in Sect.2.3. Here we test the detection of slow and fast species and the accuracy of model reduction.

4.4.1 Slowness index

The detection of slow fast species is tested by comparison with a numerical method introduced in [22]. This method consists in simulating trajectories $x_i(t)$ for each species i of the model and comparing them to the imposed trajectories x_i^* calculated as solutions of quasi-steady state equations cf. Eq. (5). Precisely, x_i^* is the solution of $\sum_j k_j S_{ij} \mathbf{x}^{\alpha_j} = 0$ in which all species of indices $l \neq i$ are replaced by their simulated values $x_l(t)$. Like in [22] we use the slowness index $I_i(t) = |\log(x_i(t)/x_i^*(t))|$. Fast species obey quasi-steady state conditions (see Eq. (5) and Sect.2.1). Therefore, for fast species, I_i is close to

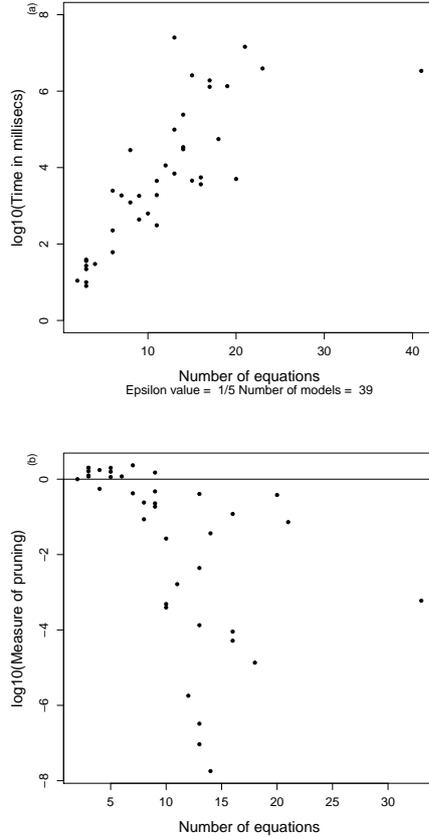


Fig. 4 (a) Semi-log plot showing \log_{10} of CPU running time (in milliseconds) versus number of equations (which may be greater than number of variables because of conservation laws) for ε value of $1/5$. (b) Plot showing the efficacy of tree pruning strategy for ε of $1/5$. The scatterplot plots the log of measure of pruning against the number of equations (which may be greater than number of variables because of conservation laws) in Biomedels database. The measure of pruning is computed as per Sect. 4.3.

zero. For slow species, the trajectories $x_i(t)$ are different from $x_i^*(t)$ and the index I_i is high. Fig. 6 shows the values of this index for all the species in the cell cycle model BIOMD00000005. In our tropical method a species is fast or slow depending how the orders $\nu_i = \mu_i - a_i$ compare to a timescale threshold. For $\varepsilon = 1/9$, we find three tropical solutions, already discussed in Sect.3.4. For the solution \mathbf{a}_3 the species 1, 2, and 5 are fast and the species 3, 4, and 6 are slow (timescales 1 min or slower). This solution leads to the Reduced model 1 described in Sect.3.4. In contrast, species 5 is slow for the two other equilibrations corresponding to the Reduced model 2. The numerical method based on the slowness index classifies species 1,2, and 5 as fast and is thus

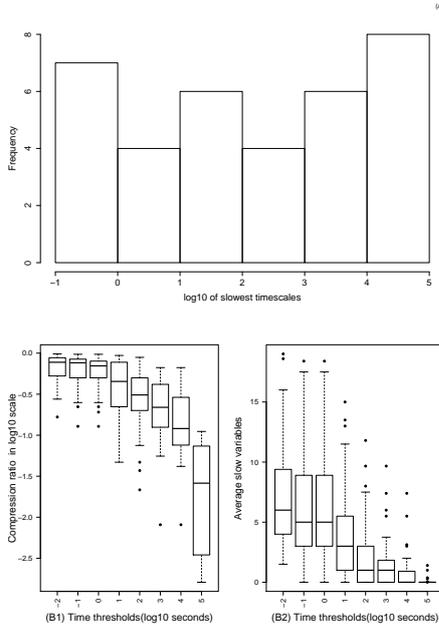


Fig. 5 (A) Histogram showing the distribution of slowest timescales for 35 models corresponding to $\varepsilon = 1/23$. The quasi buffered species (cf. Sect. 4.2) are removed before performing this step. (B1) and (B2) are boxplots showing compression ratio and average number of slow variables in the Biomodels database for different values of time threshold θ . The compression ratio is defined as the average number of slow variables / number of variables in the model. The quasi buffered species (cf. Sect. 4.2) are removed before performing this analysis. In (B1), a point represents the compression ratio over all the tropical equilibrations for each model with respect to different time thresholds. Likewise, in (B2), a point represents the average slow variables. The number of slow variables were computed based on rescaled orders (computed as in Eq. (15)) and certain time threshold in seconds (model time). The time thresholds -2 to 5 in the plot are the \log_{10} transformed values of time thresholds $0.01, 0.1, 1, 10, 100, 1000, 10000, 100000$ in secs. The boxplots correspond to value of ε of $1/23$ respectively. The boxplots of other ε look similar.

compatible with the new method for the tropical solution \mathbf{a}_3 (Fig. 6a)). The Reduced model 1 corresponding to the tropical solution \mathbf{a}_3 reproduces with good accuracy the limit cycle oscillations of the cell cycle model as shown in Fig. 6c).

4.4.2 Accuracy of the reduction

A quantitative estimate of reduction accuracy can be based on the L^2 norm of the difference between trajectories $\mathbf{x}(t)$, $\mathbf{x}_{red}(t)$ simulated with the full and reduced model, respectively. However, because periods are slightly changed by the reduction, the error could be defined as $err = \inf_a \|\mathbf{x}(t) - \mathbf{x}_{red}(at)\| / \|\mathbf{x}(t)\|$, where a is a time scaling parameter close to 1. For the trajectories shown in

Fig. 6c), err is less than 0.01 and the optimal scaling parameter is $a = 1.0002$ (the relative change of the period is 0.0002).

The two other equilibrations lead to the Reduced model 2 that is at least as accurate as the Reduced model 1 (in short, in the Reduced model 2 species 5 is considered slow and is not eliminated). This reduction accurately reproduce the dynamics not only on the limit cycle attractor, but also when initial data is far from this attractor. This is illustrated in Fig. 6d). We have simulated the full model and the two reduced models starting from several initial data \mathbf{x}_{0i} , $i = 1, \dots, 3$. The initial data of the reduced models is obtained by projection on the corresponding BIOMD00000005 invariant manifolds. For example, the reduced model 1 evolves on an invariant manifold whose equations (up to small correcting terms) are $x_5 = k_6/(k_4x_2)$, $x_1 = k_3x_2/k_1$. By computing the eigenvalues of the Jacobian of system (21) we found that this invariant manifold has an attractive, stable region (all eigenvalues, except the zero ones corresponding to exact conservation laws, have negative real parts) and an unstable region (where there are eigenvalues with positive real parts). The initial data vectors \mathbf{x}_{01} and \mathbf{x}_{02} are close to the unstable region of the invariant manifold. Therefore, trajectories starting from these initial data first get away from the manifold and after large excursions approach the attractive part of the manifold. Reduced model 2 is able to reproduce these transients but not the Reduced model 1 (Fig. 6d) because the latter is valid only on the attractive invariant manifold.

4.5 Comparison with COPASI time separation method

We compared our proposed method against the existing tool COPASI [13, 32]. COPASI proposes a modified ILDM (intrinsic low dimensional manifold) method to perform time separation of the variables. This method computes slow and fast modes which are transformations of species concentrations as described in [4, 35, 32]. More precisely, COPASI performs a Schur block decomposition of the Jacobian matrix \mathbf{J} , consisting in finding a non-singular matrix T such that $T^{-1}JT = \begin{pmatrix} S_{slow} & 0 \\ 0 & S_{fast} \end{pmatrix}$, where S_{slow}, S_{fast} have real Schur form, i.e. they are upper triangular matrices with possibly non-vanishing elements on the first subdiagonal. The time threshold (needed to separate the slow and fast blocks of the Jacobian matrix) is automatically captured in this method by finding a gap in the spectrum of the Jacobian (cf. Sect.2.1).

In order to compare the modified ILDM method against our tropical and slowness index methods, we computed the fast space of the model using COPASI for 100 time steps between 1 and 100 min and checked the contribution of each species to this fast space. COPASI defines the contribution of a species i to a mode j as the matrix element T_{ji}^{-1} . These contributions of various species to fast modes are provided by COPASI as fractions p_i , where i is for the species, $0 \leq p_i \leq 1$, $\sum_i p_i = 1$. COPASI declare species with largest contribution to the fast space (largest p_i) as fast species. For exactly the same trajectory,

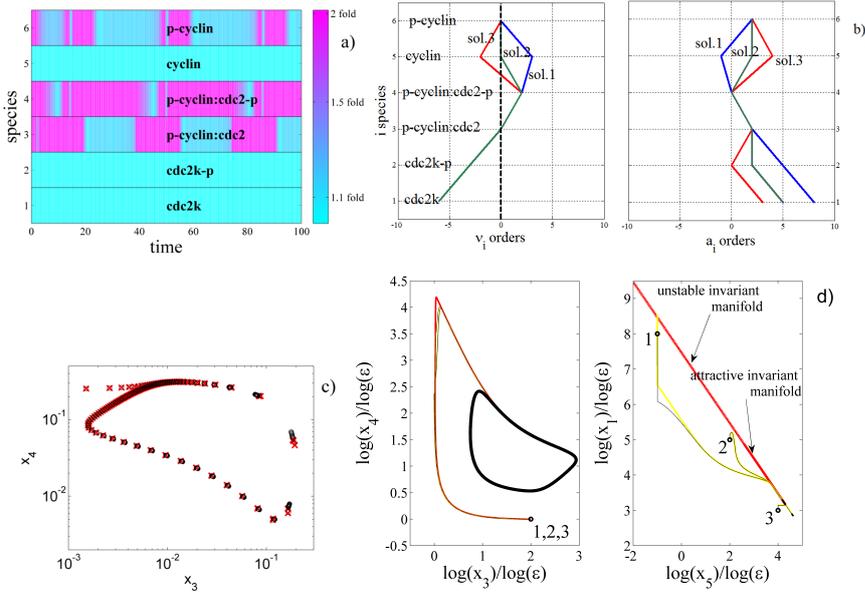


Fig. 6 Testing tropical slow/fast decomposition and accuracy of reduction of BIOMD00000005. a) the slowness index is represented as a function of time on trajectories: slow variables have large index (fold ratio); b) left : the orders $\nu_i = \mu_i - a_i$ are represented for different species and for three tropical solutions; slow species have orders above a threshold corresponding to 1 min and represented as a dotted line; right : orders a_i for different species and tropical solutions give species concentrations; the higher a_i , the lower the concentration ; c) comparison of the limit cycle trajectories computed with the full (black circles) and reduced model (red crosses); d) Model trajectories for the full model (black), reduced model 1 (red) and reduced model 2 (yellow), starting from three initial data, corresponding to three different tropical equilibrations. The limit cycle attractor is contained in an invariant manifold. The reduced model 1 provides a good approximation of the dynamics on the invariant manifold (such as starting from initial data 3), but not outside. The reduced model 2 is accurate also outside the invariant manifold (see trajectories starting from equilibrations 1 and 2).

we have computed the values of our slowness indices $s_i = |\log(x_i(t)/x_i^*(t))|$. For fast species, s_i should be close to zero. Fig. 6a) and b) summarizes the comparison between the slowness index and the tropical method. The tropical solution \mathbf{a}_3 leads to the Reduced model 1 (cf. Sect.3.4) and copes with the limit cycle trajectory used in this test. The timescale orders ν_i of the variables for this tropical solutions identify species x_1, x_2, x_5 as fast and species x_3, x_4, x_6 as slow (see also Sect.3.4). As can be seen in Fig. 6a) the slowness index of species x_1, x_2, x_5 is close to zero for all times. The slowness index of species x_3, x_4, x_6 can reach large values. Therefore the tropical method and the slowness index method provide exactly the same timescale decomposition. COPASI time separation can not be compared directly to the tropical method, because it generates a timescale decomposition that changes with time and which is valid for a trajectory. However, it can be compared with

the slowness index decomposition. Fig. 7 summarizes the comparison between COPASI and the slowness index. It should be noted that the species x_3 is automatically eliminated by COPASI using the single conservation law present in the model. The slowness index and COPASI contribution to fast space should be anticorrelated: when the first one is small the latter should be big and vice versa. Species x_1 and x_5 have high contribution towards the fast space and very low slowness index. For these species we can say there is consistence between COPASI and slowness index. Species x_2 also has large contribution to fast space except for some intervals where COPASI may classify it as slow. Our method unambiguously classifies this species as fast (its slowness index is very low for all times). Most importantly, COPASI fails to identify correctly time intervals where species 6 is slow as indicated by the large value of the slowness index co-existing with large values of contribution p_i (as large as for species 1 and 2). According to COPASI this species is similar to fast species x_2 , whereas our methods indicate it to be similar to the slow species x_4 and x_3 . As a matter of fact, COPASI determines slow variables by comparing the values of contributions p_i to the fast and slow modes. In spite of the spectral gap, the differences of the indices p_i between species that are considered slow and fast can be relatively small and therefore the classification is not robust. In contrast, our methods classify species in a robust way. Indeed, we directly associate timescales to species, via the orders ν_i and these timescales are well separated for slow and fast species. Using these orders, we found that the fastest slow species x_3 and x_6 are 100 times slower than the slower fast species x_5 . Furthermore, as shown in Fig.6a, our slowness index is very sensitive to differences in timescales. Fast species x_1 , x_2 , x_5 keep this index low for all times, whereas the corresponding COPASI indices are not always high. Generally, it should not be recommended to use species contributions to modes as indicative of their timescales, as COPASI does. For instance, the sum of two or more species can be a slow mode, even if all these species are fast (cf. Sect. 2.3, this situation is typical for fast cycles). The fast species have in this case high contributions to a slow mode which may qualify them as slow according to the contribution criterion.

5 Discussion

We provide an algorithm for reduction of biochemical networks. The method is based on computation of tropical equilibrations. Tropical equilibrations generalize the notion of steady state and correspond to compensation of dominant reaction fluxes and to slow dynamics on invariant manifolds under the action of weaker fluxes. We have proposed an algorithm to compute tropical equilibrations. We have used this algorithm for computing the fast/slow partition of chemical species that is the first and often critical step in model reduction algorithms. In particular, the number of slow species provides the size of minimal dynamic models to which complex biochemical networks can be reduced. The validity of our reductions depends on concentration orders. Such

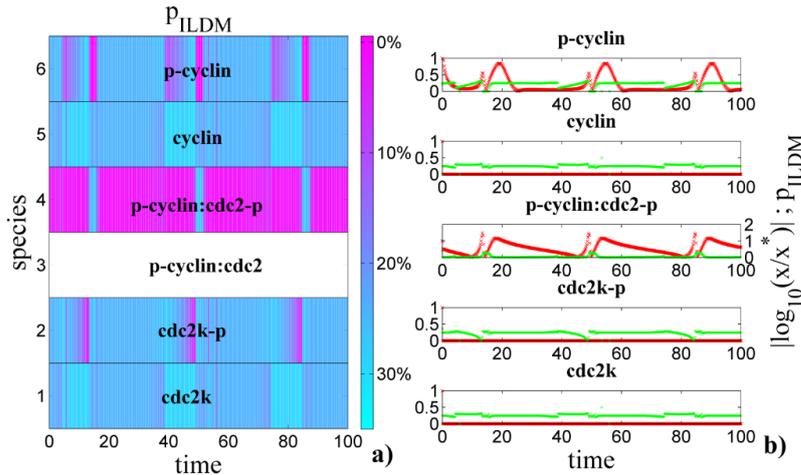


Fig. 7 Summary of the analysis of model BIOMD0000005 using ILDM method and comparison with our method based on the slowness index. The model was simulated in COPASI for 400 seconds with interval size of 0.01 and default initial concentrations. The species 3 is eliminated using the single conservation law. For all the remaining species we represent the time dependence of their contributions p_i , where $0 \leq p_i \leq 1$, $\sum_i p_i = 1$ to the fast space. The fractions p_i are generated by COPASI and the values p_i are color coded in the left panel a). Chemical species with largest contribution to fast space (largest p_i) are supposed to be fast species (as explained in [32]). Therefore p_i (in green in right panel b)) and our slowness index $s_i = |\log(x_i/x_i^*)|$ (in red) should be anti-correlated. This is well verified for species 1, 4, 5 (p is relatively high when s is close to zero, and close to zero when s is relatively high, but it is not well verified for species 6 (p-cyclin)).

conditions correspond to large domains of the phase space, therefore to robust reductions [22].

Our method facilitates analysis of complex models. Reductions emphasize important and sensible parameters and unravel essential mechanisms. Furthermore, lowering the number of parameters and variables can allow the applicability of many existing formal methods, that do not work on large models but may work on reduced models.

The benchmarking of our algorithm on the Biomodels database shows that a significant dimension compression can be performed on cell dynamics models at timescales of 1000s and larger. Starting with complex models having more than 30 variables, minimal models have median numbers of 2 slow variables. This suggests that, at least piece-wisely in parameter and phase space, the tasks fulfilled by molecular networks are relatively simple. The need for having complex machineries with many regulators to perform simple tasks (such as relaxation to steady states or limit cycle oscillations) are justified by system robustness. A system having a very large number of variables and parameters, but only a few independent degrees of freedom is generically robust with respect to perturbations of variables and parameters [8].

Solving the tropical equilibration problem and finding a slow-fast decomposition is the first step for model reduction. The remaining steps consist in elimination of the fast variables by solving systems of algebraic equations. We have shown how this can be performed for a simple example. In the case of more complex models, the elimination can be performed numerically, or symbolically. Tropical methods can simplify this task by replacing the full systems by tropically truncated systems. In particular, the binomial or toric case when the truncation has only two monomials is particularly interesting because for this case there are rapid algorithms for computing steady states [18]. Higher approximation can be provided by Newton-Puiseux expansions [24], that encompass tropical solutions in their lowest order. Although the calculations needed for formal reduction could be long, once the model is reduced, it can be used in various applications, such as a part of larger networks, or in models of tissues and organisms where the same biochemical network has to be replicated in several interacting cells. Furthermore, our reductions have a strong geometrical basis. In future work, we will exploit this property to show how to endow the reduced model with a reaction network structure and how to identify inclusion relations among different reduced models. We will also investigate the formal dependence of the set of tropical equilibrations on the parameters. This would allow us to formally classify possible reductions and phase portraits of a model with a given topology and reaction rates.

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